

## Inventory of the fish fauna from Laranjinha River, Paranapanema River system, Brazil

Bruno Ambrožio Galindo<sup>1</sup> , Renata Rúbia Ota<sup>2</sup> , Thiago Deruza Garcia<sup>3\*</sup> , Raul Henrique Cardoso Nascimento<sup>3</sup>, Willian Massaharu Ohara<sup>4</sup>, Augusto Seawright Zanatta<sup>1</sup>, Dhiego Gomes Ferreira<sup>1</sup>, Caroline Apolinário-Silva<sup>5</sup>, Wilson Frantine-Silva<sup>6</sup> , Sandremir de Carvalho<sup>7</sup>, Alexandre Derly Augusto Costa<sup>3</sup>, Silvia Helena Sofia<sup>5</sup> & Oscar Akio Shibatta<sup>8</sup> 

<sup>1</sup>Universidade Estadual do Norte do Paraná, Ciências Biológicas, Cornélio Procópio, PR, Brasil.

<sup>2</sup>Universidade Estadual de Maringá, Coleção Ictiológica do Nupélia, Maringá, PR, Brasil.

<sup>3</sup>Universidade Estadual de Londrina, Programa de Pós-graduação em Ciências Biológicas, Londrina, PR, Brasil

<sup>4</sup>Universidade Federal de Rondônia, Engenharia de Pesca, Presidente Médici, RO, Brasil.

<sup>5</sup>Universidade Estadual de Londrina, Departamento de Biologia Geral, Londrina, PR, Brasil.

<sup>6</sup>Universidade Estadual do Norte Fluminense Darcy Ribeiro, Laboratório de Ciências Ambientais, Campos dos Goytacazes, RJ, Brasil.

<sup>7</sup>Universidade Estadual do Norte do Paraná, Ciências Biológicas, Bandeirantes, PR, Brasil.

<sup>8</sup>Universidade Estadual de Londrina, Biologia Animal e Vegetal, Londrina, PR, Brasil.

\*Corresponding author: Thiago Deruza Garcia, e-mail: [thiago.duruza@hotmail.com](mailto:thiago.duruza@hotmail.com)

GALINDO, B.A., OTA, R.R., GARCIA, T.D. , NASCIMENTO, R.H.C., OHARA, W.M., ZANATTA, A.S., FERREIRA, D.G., APOLINÁRIO-SILVA, C., FRANTINE-SILVA, W., CARVALHO, S., COSTA, A.D.A., SOFIA, S.H., SHIBATTA, O.A. **Inventory of the fish fauna from Laranjinha River, Paranapanema River system, Brazil.** Biota Neotropica 20(2): e20200962. <https://doi.org/10.1590/1676-0611-BN-2020-0962>

**Abstract:** This work is the most comprehensive survey of the Laranjinha River's fishes, a tributary of the Cinzas River, Paranapanema River basin. Throughout its course, there is only a low-height dam, including a transposition system located 98 km from its mouth. The sampling was carried out in nine locations, from the source to the mouth, with six field incursions in each location, using different fishing gear. A total of 11,924 fish were collected, distributed in seven orders, 27 families, and 100 species. The most representative order in the number of species was Siluriformes, followed by Characiformes. As for the families, Loricariidae comprised 21% and Characidae 14% of species richness. *Phalloceros harpagos* was the species with the highest absolute abundance, representing 11.3% of the total, followed by *Hypostomus ancistroides* with 9.8%. However, considering the average abundance and frequency of occurrence, *Hypostomus ancistroides* was the most abundant species, followed by *Hypostomus cf. paulinus*, *Psalidodon aff. paranae* and *Phalloceros harpagos*. Among the collected species, the *Apteronotus acidops*, *Brycon orbygnianus*, *Brycon nattereri*, *Crenicichla jupiaensis*, and *Rhinelepis aspera* were classified as endangered on the most recent IUCN Red List. Also, from the total sampled fish, 9.8% are considered non-native species. Among the native species recorded, 10 species are large migratory species, which indicates that the Laranjinha River is a route for spawning and maintenance of species diversity in the middle Paranapanema River. Therefore, the Laranjinha River is a heritage of fish diversity and deserves special attention in its preservation.

**Keywords:** Checklist; fish diversity; freshwater; upper Paraná River.

## Inventário da ictiofauna do rio Laranjinha, sistema do rio Paranapanema, Brasil

**Resumo:** Este é o levantamento mais abrangente de peixes do rio Laranjinha, um afluente do rio das Cinzas, bacia do rio Paranapanema. Ao longo de sua rota, existe apenas uma pequena barragem com um sistema de transposição localizado 98 km de sua nascente. A amostragem foi realizada em nove locais, desde a nascente até a foz, com seis incursões de campo em cada local, com o auxílio de diferentes artes de pesca. Foram coletados 11.924 indivíduos, distribuídos em sete ordens, 27 famílias e 100 espécies. A ordem mais representativa foi Siluriformes, seguida por Characiformes. Quanto às famílias, Loricariidae compôs 21% e Characidae 14% da riqueza de espécies. *Phalloceros harpagos* foi a espécie com maior abundância absoluta, representando 11,3% do total, seguida por *Hypostomus ancistroides*, com 9,8%. No entanto, considerando a abundância média e a frequência de ocorrência, *Hypostomus ancistroides* foi a espécie mais abundante, seguida por *Hypostomus cf. paulinus*, *Psalidodon aff. paranae* e *Phalloceros harpagos*. Entre as espécies coletadas, *Apteronotus acidops*, *Brycon orbygnianus*, *Brycon nattereri*, *Crenicichla jupiaensis* e *Rhinelepis aspera* estão listadas em categorias de ameaça na Lista Vermelha da IUCN mais recente.

Além disso, do total de indivíduos amostrados, 9,8% são considerados espécies não nativas. Entre as espécies nativas registradas, 10 espécies são migratórias de grande porte, o que indica que o rio Laranjinha é uma rota de desova e manutenção da diversidade de espécies no médio rio Paranapanema. Portanto, o rio Laranjinha é um patrimônio da diversidade de peixes e merece atenção especial em sua preservação.

**Palavras-chave:** Água doce; alto rio Paraná; diversidade de peixes; lista de espécies.

## Introduction

In the last few decades, human actions have caused numerous environmental changes, which emphasizes the need to know the local biodiversity, in order to reduce harmful activities. To this extend, the creation of inventories contribute to the discovery and description of new species before their extinction, also contributing to the creation of new records, to the knowledge about geographical distribution, the documentation of non-native species, the definition of biogeographic patterns, and the establishment of suitable conservation strategies (Costello et al. 2011, Ota et al. 2015, Frota et al. 2019).

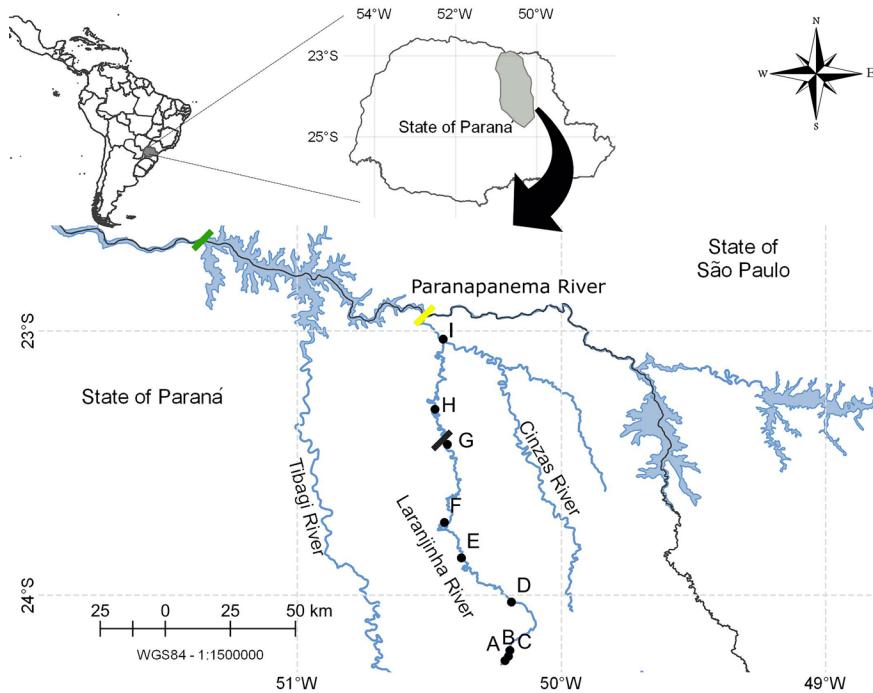
The fishes of the upper Paraná River basin are the most studied in Brazil. The basin has more than 310 fish species (Langeani et al. 2007), a number currently outdated, considering new records and the recent discovery of new species (e.g. Frota et al. 2016, Froehlich et al. 2017, Cavalli et al. 2018, Ota et al. 2018, Jarduli et al. 2020). The Paranapanema River, a major tributary of the upper Paraná River, is currently fragmented by several hydroelectric dams (Britto & Carvalho 2006), and therefore, its dam-free tributaries are of great importance for the maintenance of fish species (Hoffmann et al. 2005, Silva et al. 2017, Galindo et al. 2019, Lansac-Tôha et al. 2019). Besides, the Cinzas River is an important tributary of the Paranapanema River basin, which is the main watercourse of the region known as “Norte Pioneiro” (Pioneer North) of the Paraná state, and it is essential for maintaining ichthyofauna biodiversity of the Capivara reservoir (Vianna & Nogueira 2008, Orsi 2010).

Fish inventories were carried out in the Paranapanema River tributaries, including the Cinzas, Jaguariaíva and Tibagi rivers (e.g., Shibatta et al. 2002, Castro et al. 2003, Cetra et al. 2012, Cionek et al. 2012, Costa et al. 2013, Hoffmann et al. 2005, Cetra et al. 2016, Frantine-Silva et al. 2015, Almeida et al. 2018, Jerep & Shibatta 2017, Claro-García et al. 2018, Frota et al. 2020, Jarduli et al. 2020, Garcia et al. 2020). However, a comprehensive checklist of the Laranjinha River ichthyofauna has not been published yet, except for a small streams (e.g., Costa et al. 2013), and the scientific knowledge of its fish species remains unknown. Furthermore, projects of Small Hydropower Plants (SHPs) are being developed, which represents a potential threat to the species of this river (Galindo et al. 2019). Thus, this study aimed to provide the first inventory of the Laranjinha River fish fauna.

## Material and Methods

### 1. Study area

The Laranjinha River is entirely situated in the northeastern portion of the state of Paraná, and it is the main tributary of the left bank of the Cinzas River, which pours to the left bank of the middle Paranapanema River (Figure 1 and 2). The headwater is in the municipality of Ventania ( $24^{\circ}14'43.43''S$ ;  $50^{\circ}14'32.78''W$ ), at 984 m of altitude, and the mouth is situated 4 km from the town of Itambaracá ( $23^{\circ}01'03.51''S$ ;  $50^{\circ}24'22.68''W$ ), at 348 m of altitude.



**Figure 1.** Map of the points sampled along the Laranjinha river (black circles). The green rectangle represents the Capivara reservoir, and the yellow rectangle represents the Canoas I reservoir, both on the Paranapanema River basin.



**Figure 2.** Partial view of collection points along the Laranjinha River, illustrating the environments and marginal vegetation. Points near source (a-c), points in the middle portion (d-f), and places near the mouth (g-i).

The Laranjinha River, with its meanders, extends through 350 km long, and it is situated in the sedimentary basin of the state of Paraná, with its source and mouth in the second and third plateaus, respectively (Santos et al. 2006). Between the years 1956 and 1960, a Small Hydropower Plant (SHP) was built 98 km from the mouth of the river. This dam never went into operation, and in 2006, a fish pass system was built in it (Schwartz 2006; Figure 2G).

## 2. Data collection

Nine sites for sampling were distributed from the source to the mouth of the Laranjinha River (Figures 1 and 2; Table 1). Six collections were carried out in each site, with the aid of different fish gears (seines, gill nets, cast nets and sieves), from October 2010 to April 2012. The collections were authorized by SISBIO (Sistema de Autorização e Informação em Biodiversidade), Ministério do Meio Ambiente, under the nº 23315-1.

After the capture of the fish, the specimens were anesthetized with 10% benzocaine. This substance promotes a reduction in gill ventilation due to the depression of spinal respiratory centers, promoting a decrease in blood flow through the gills (Mattson & Riple 1989; Tytler & Hawkins 1981). After performing the opercular movements, the fish were fixed in 10% formalin and preserved in 70% ethanol in the Laboratório de Genética e Conservação of the Universidade Estadual do Norte do Paraná (GECON/UENP-CCP) and identified in species-level. The validity of the species was checked using Fricke et al. (2019), and the classification was based using Van der Laan et al. (2020). The species were identified following Ota et al. (2018), Jarduli et al. (2020), Terán et al. (2020), and in the lack of information about the species from the authors before mentioned, personal communication with specialists took place (i.e., FC Jerep, CDCM de Santana, CAM Oliveira and CH Zawadzki).

The record of *Poecilia reticulata* Peters 1859 and *Imparfinis schubarti* (Gomes 1956) were based on Costa et al. (2013). Non-native species were based in Orsi & Agostinho (1999), Lobón-Cerviá & Bennemann (2000), Langeani et al. (2007), Júlio-Júnior et al. (2009), Britton & Orsi (2012), Ortega et al. (2015), Azevedo-Santos et al. (2016), Ota et al. (2018), Pelicice et al. (2018) and Jarduli et al. (2020) (Table 2). Vouchers of all species were deposited in the Museu de Zoologia da Universidade Estadual de Londrina (MZUEL). The abundances of orders, families, and species were

conducted using the Statistica 7.0 software (StatSoft Inc. 2011). The Kendeigh index of abundance (1944) of each species was calculated as  $KI = \sqrt{FO \cdot M}$ ; where  $KI$  is the abundance index, and  $FO$  is the frequency of occurrence calculated as the number of sites where the species were captured, and divided by the total number of sites multiplied by 100, and  $M$  is the mean number of specimens of each species. The classification of species was done by sorting the values in descending order. The dominance index and evenness were calculated with the program PAST v. 2.17c (Hammer et al. 2001).

**Table 1.** Description of the sampled sites in the Laranjinha river, a tributary of the Paranapanema River, Paraná. “Localities from which coordinates were not originally georeferenced were determined through Google Earth, and are approximate.”

Site	Locality	City	Coordinates	Environment Description
A	Pedreira	Ventania	24°15'8.55"S 50°12'1.35"W	The riparian vegetation is well-preserved on both river banks; substrate predominantly sandy and slabs; average width of approximately 4m. Near pastureland and agriculture areas.
B	Cachoeira	Ventania	24°14'37.57"S 50°12'24.48"W	The riparian vegetation is well-preserved on both river banks; substrate predominantly rocks and sand; average width of approximately 4m. Eight kilometers above receives a massive load of domestic effluents.
C	SANEPAR	Ventania	24°13'24.56"S 50°11'57.88"W	The riparian vegetation is well-preserved on the right river bank, a predominance of grass in the left river bank; mostly sedimentary and clayey substrate; average width of approximately 5m; great anthropic disturbance by the presence of a bridge and rural road.
D	Distrito da Moquém	Ventania	24°01'36.6"S 50°11'21.6"W	The riparian vegetation is well-preserved on both river banks (showing the best conditions among all stretches studied); some rocky walls, and predominantly sandy bottom; average width of approximately 13m. Stretch including a lake between a rapid and a small waterfall. It is inserted in a private property for exotic tree planting for logging.
E	Figueira	Figueira	23°51'34.73"S 50°22'43.04"W	The riparian vegetation is absent on both river banks (mostly grass); mainly rocky and sandy substrate; average width of approximately 14m. It is located in pastureland and agriculture areas, upstream of a thermal power plant.
F	Ibaiti	Ibaiti	23°43'31.88"S 50°26'34.42"W	The riparian vegetation is reduced on the left bank (significantly modified by agricultural activity) while the right bank is well-preserved; average width of approximately 40m; some rapids and predominantly rocky and sandy substrate. It is located upstream of a sugar and ethanol plant and surrounded by agriculture areas.
G	Barragem da Corredeira	Ribeirão do Pinhal	23°17'49.95"S 50°28'43.27"W	Riparian vegetation is present on both river banks, well-preserved, alternating sections with grass predominance stretches; includes a dam separating an upstream lake (average width of approximately 63m) and downstream stretch including rapids and some deeper wells (average width of 48m); sand and clayey substrate in the lake and a rocky substrate along downstream stretch; surrounded by pastureland and agriculture areas.
H	Santa Amélia	Santa Amélia	23°24'53.06"S 50°27'8.60"W	The riparian vegetation is poorly preserved on both river banks; large rapids and some deeper wells; predominantly rock substrate; average width of approximately 55m; surrounded by pastureland and agriculture areas.
I	Foz	Bandeirantes	23°1'53.90 S 50°26'51.90"W	The riparian vegetation is poorly preserved on both river banks; some upstream rapids, however, a calm water segment prevails until the confluence between Laranjinha and Cinzas Rivers, including a rocky and sandy (primary) substrate; average width of approximately 48m. It is surrounded by pastureland and agriculture areas.

## Ichthyofauna of the Laranjinha River basin

**Table 2.** List of species captured along the Laranjinha River (sites A to I, see Figures 1 and 2, and Table 1 for location and characterization), Paraná, Brazil, and their respective catch abundance. ♣ Non-native species, according to, Langeani et al. (2007), Júlio-Júnior et al. (2009), Ota et al. (2018), Jarduli et al. (2020) and Orsi personal communication. ♦ Additional species recorded in the basin by Costa et al. (2013).

Táxon	Sampled Sections									Total	FO	FO%	Mean	FO%*Mean	KI	Voucher											
	A	B	C	D	E	F	G	H	I																		
<b>CYPRINIFORMES</b>																											
<b>Cyprinidae</b>																											
<i>Cyprinus carpio</i> Linnaeus 1758♣										2		2	1.0	11.1	0.2	2.46	1.6 MZUEL 10559										
<b>CHARACIFORMES</b>																											
<b>Crenuchidae</b>																											
<i>Characidium aff. zebra</i>										4	19	1		24	3.0	33.3	2.7	88.88	9.4 MZUEL 09350								
<b>Erythrinidae</b>																											
<i>Hoplias gr. malabaricus</i>										1	6	17	62	6	2	29	123	7.0	77.8	13.7	1062.96	32.6 MZUEL 19806					
<b>Parodontidae</b>																											
<i>Apareiodon ibitiensis</i> Campos 1944										1	4		1	10		2	18	5.0	55.6	2.0	111.11	10.5 MZUEL 16063					
<i>Apareiodon piracicabae</i> (Eigenmann 1907)										35	45	44	91	79	143		437	6.0	66.7	48.6	3237.03	56.9 MZUEL 09257					
<i>Parodon nasus</i> Kner 1859																	27	2.0	22.2	3.0	66.66	8.2 MZUEL 09365					
<b>Serrasalmidae</b>																											
<i>Metynnis lippincottianus</i> (Cope 1870) ♣																	1	6	7	2.0	22.2	0.8	17.28	4.2 MZUEL 09519			
<i>Piaractus mesopotamicus</i> (Holmberg 1887)																	3	6	9	2.0	22.2	1.0	22.22	4.7 MZUEL 09471			
<i>Serrasalmus maculatus</i> Kner 1858																	32	1	14	47	3.0	33.3	5.2	174.07	13.2 MZUEL 16059		
<b>Anostomidae</b>																											
<i>Leporellus vittatus</i> (Valenciennes 1850)																	2	2	4	2.0	22.2	0.4	9.87	3.1 MZUEL 09556			
<i>Leporinus amblyrhynchus</i> Garavello & Britski 1987										20	23	9	43	11	15		121	6.0	66.7	13.4	896.29	29.9 MZUEL 09262					
<i>Leporinus friderici</i> (Bloch 1794)																	3	9	55	3.0	33.3	6.1	203.70	14.3 MZUEL 09455			
<i>Leporinus octofasciatus</i> Steindachner 1915										5	3	12	10			16	46	5.0	55.6	5.1	283.95	16.9 MZUEL 09440					
<i>Leporinus striatus</i> Kner 1858																	15	17	42	3.0	33.3	4.7	155.55	12.5 MZUEL 09520			
<i>Megaleporinus obtusidens</i> (Valenciennes 1837)																	10	1	18	3.0	33.3	2.0	66.66	8.2 MZUEL 09431			
<i>Schizodon borellii</i> (Boulenger 1900)										4	1		1	79			85	4.0	44.4	9.4	419.75	20.5 MZUEL 09467					
<i>Schizodon nasutus</i> Kner 1858										25	29	49	2	16			121	5.0	55.6	13.4	746.91	27.3 MZUEL 09356					
<b>Curimatidae</b>																											
<i>Cyphocharax modestus</i> (Fernández-Yépez 1948)										27	4	75	1	1	17		125	6.0	66.7	13.9	925.92	30.4 MZUEL 09448					
<i>Steindachnerina insculpta</i> (Fernández-Yépez 1948)										2		12	109	12	37		172	5.0	55.6	19.1	1061.72	32.6 MZUEL 09451					
<b>Prochilodontidae</b>																											
<i>Prochilodus lineatus</i> (Valenciennes 1837)																	120	40	29	4	4	197	5.0	55.6	21.9	1216.04	34.9 MZUEL 09437
<b>Triportheidae</b>																											
<i>Triportheus nematurus</i> (Kner 1858) ♣																	1	43	44	2.0	22.2	4.9	108.64	10.4 MZUEL 09458			

continue...

...continue

Táxon	Sampled Sections									Total	FO	FO%	Mean	FO%*Mean	KI	Voucher
	A	B	C	D	E	F	G	H	I							
<b>Bryconidae</b>																
Bryconinae																
<i>Brycon nattereri</i> Günther 1864		82	5							87	2.0	22.2	9.7	214.81	14.7	MZUEL 09470
<i>Brycon orbignianus</i> (Valenciennes 1850)					50					50	1.0	11.1	5.6	61.728	7.9	MZUEL 09485
Salmininae										0						
<i>Salminus brasiliensis</i> (Cuvier 1816)						1				1	1.0	11.1	0.1	1.23	1.1	MZUEL 09527
<i>Salminus hilarii</i> Valenciennes 1850						1				1	1.0	11.1	0.1	1.23	1.1	MZUEL 09518
<b>Acestrorhynchidae</b>																
<i>Acestrorhynchus lacustris</i> (Lütken 1875)				56	38	61				155	3.0	33.3	17.2	574.07	24.0	MZUEL 09543
<b>Characidae</b>																
Stethaprioninae																
<i>Astyanax lacustris</i> (Lütken 1875)	1	18	42	96	242	106	134			639	7.0	77.8	71.0	5522.22	74.3	MZUEL 09538
<i>Hyphessobrycon eques</i> (Steindachner 1882) ♀								12		12	1.0	11.1	1.3	14.81	3.8	MZUEL 09464
<i>Moenkhausia</i> aff. <i>intermedia</i>								157		157	1.0	11.1	17.4	193.82	13.9	MZUEL 09457
<i>Oligosarcus paranensis</i> Menezes & Géry 1983	45	13	18							76	3.0	33.3	8.4	281.48	16.8	MZUEL 09534
<i>Psalidodon bockmanni</i> (Vari & Castro 2007)	1	36	70	7	4	21	17			156	7.0	77.8	17.3	1348.14	36.7	MZUEL 09354
<i>Psalidodon</i> aff. <i>fasciatus</i>		201	9	11	86	11	14			332	6.0	66.7	36.9	2459.25	49.6	MZUEL 09541
<i>Psalidodon</i> aff. <i>paranae</i>	571	134	70	14	9	22	2	1		823	8.0	88.9	91.4	8128.39	90.2	MZUEL 09441
Characinae																
<i>Galeocharax gulo</i> (Cope 1870)						118	42	43		203	3.0	33.3	22.6	751.85	27.4	MZUEL 09259
Aphyocharacinae																
<i>Aphyocharax cf. dentatus</i> Eigenmann & Kennedy 1903 ♀								1		1	1.0	11.1	0.1	1.23	1.1	MZUEL 09463
Cheirodontinae																
<i>Odontostilbe weitzmani</i> Chuctaya, Bührnheim & Malabarba 2018			7	3						10	2.0	22.2	1.1	24.69	5.0	MZUEL 09557
<i>Serrapinnus notomelas</i> (Eigenmann 1915)		26	48	2						76	3.0	33.3	8.4	281.48	16.8	MZUEL 09435
Stevardiinae																
<i>Bryconamericus</i> aff. <i>iheringii</i> (Boulenger 1887)	13	112	19	18	98					260	5.0	55.6	28.9	1604.93	40.1	MZUEL 09528
<i>Piabarchus</i> aff. <i>stramineus</i>		2	4	2	15					23	4.0	44.4	2.6	113.58	10.7	MZUEL 09433
<i>Piabina argentea</i> Reinhardt 1867				2	5					7	2.0	22.2	0.8	17.28	4.2	MZUEL 09348
<b>GYMNOTIFORMES</b>																
<b>Apteronotidae</b>																
<i>Apteronotus acidops</i> Triques 2011						1				1	1.0	11.1	0.1	1.23	1.1	
<i>Apteronotus</i> aff. <i>albifrons</i> (Linnaeus 1766)						1				1	1.0	11.1	0.1	1.23	1.1	MZUEL 09558
<i>Apteronotus</i> cf. <i>caudimaculosus</i> Santana 2003 ♀					3					3	1.0	11.1	0.3	3.70	1.9	MZUEL 09538

continue...

## Ichthyofauna of the Laranjinha River basin

...continue

Táxon	Sampled Sections									Total	FO	FO%	Mean	FO%*Mean	KI	Voucher	
	A	B	C	D	E	F	G	H	I								
<b>Sternopygidae</b>																	
<i>Eigenmannia</i> sp.					3	2	4	3	11	23	5.0	55.6	2.6	141.97	11.9	MZUEL 09552	
<i>Sternopygus macrurus</i> (Bloch & Schneider 1801)						1	5	3		9	3.0	33.3	1.0	33.33	5.8	MZUEL 09454	
<b>Gymnotidae</b>																	
<i>Gymnotus sylvius</i> Albert & Fernandes-Matioli 1999			2		1	10	9	4	7	33	6.0	66.7	3.7	244.44	15.6	MZUEL 09546	
<i>Gymnotus omarorum</i> Richer-de-Forges, Crampton & Albert 2009						10				10	1.0	11.1	1.1	12.34	3.5	MZUEL 09517	
<b>SILURIFORMES</b>																	
<b>Trichomycteridae</b>																	
<i>Cambeva diabola</i> (Bockmann, Casatti & de Pinna 2004)	26	169	8	9	3					215	5.0	55.6	23.9	1327.16	36.4	MZUEL 09516	
<b>Callichthyidae</b>																	
<i>Callichthys callichthys</i> (Linnaeus 1758)						1				1	1.0	11.1	0.1	1.23	1.1	MZUEL 09535	
<i>Corydoras aeneus</i> (Gill 1858)							1			1	1.0	11.1	0.1	1.23	1.1	MZUEL 09547	
<i>Hoplosternum littorale</i> (Hancock 1828)						3				3	1.0	11.1	0.3	3.70	1.9	MZUEL 09544	
<b>Loricariidae</b>																	
Rhinelepininae																	
<i>Rhinelepis aspera</i> Spix & Agassiz 1829								12		12	1.0	11.1	1.3	14.81	3.8	MZUEL 09549	
Loricariinae																	
<i>Loricariichthys platymetopon</i> Isbrücker & Nijssen 1979 ♀					10	38				48	2.0	22.2	5.3	118.51	10.9	MZUEL 09460	
<i>Proloricaria prolixa</i> Isbrücker & Nijssen 1978						22	15			37	2.0	22.2	4.1	91.35	9.6	MZUEL 09525	
<i>Rineloricaria latirostris</i> (Boulenger 1900)				3	2	2		1		8	4.0	44.4	0.9	39.50	6.3	MZUEL 09530	
<i>Rineloricaria pentamaculata</i> Langeani & de Araujo 1994					1			1	2	4	3.0	33.3	0.4	14.81	3.8	MZUEL 09545	
Hypoptopomatinae																	
<i>Curculionichthys insperatus</i> (Britski & Garavello 2003)						45				45	1.0	11.1	5.0	55.55	7.5	MZUEL 09469	
<i>Neoplecostomus yapo</i> Zawadzki. Pavanelli & Langeani 2008	35	29	5	2						71	4.0	44.4	7.9	350.61	18.7	MZUEL 09542	
<i>Otothyropsis biamnicus</i> Calegari, Lehmann A. & Reis 2013				23	2					25	2.0	22.2	2.8	61.72	7.9	MZUEL 09071	
Hypostominae																	
<i>Ancistrus</i> cf. <i>cirrhosus</i> (Valenciennes 1836)						4	3	2		9	3.0	33.3	1.0	33.33	5.8	MZUEL 09522	
<i>Hypostomus albopunctatus</i> (Regan 1908)				11	1	88	19			119	4.0	44.4	13.2	587.65	24.2	MZUEL 09362	
<i>Hypostomus ancistroides</i> (Ihering 1911)	299	305	232	140	113	30	51			1170	7.0	77.8	130.0	10111.11	100.6	MZUEL 09450	
<i>Hypostomus</i> cf. <i>paulinus</i> (Ihering 1905)				6	246	91	86	194	244	1008	7.0	77.8	112.0	8711.11	93.3	MZUEL 09480	
<i>Hypostomus</i> cf. <i>topavae</i> (Godoy 1969)				1	1	1	40	32	74	205	7.0	77.8	22.8	1771.60	42.1	MZUEL 09355	
<i>Hypostomus hermanni</i> (Ihering 1905)						73	166	162		401	3.0	33.3	44.6	1485.18	38.5	MZUEL 09363	
<i>Hypostomus iheringii</i> (Regan 1908)						2		13		15	2.0	22.2	1.7	37.03	6.1	MZUEL 09473	
<i>Hypostomus nigromaculatus</i> (Schubart 1964)				29	5	6	9	9	7	65	6.0	66.7	7.2	481.48	21.9	MZUEL 09361	
<i>Hypostomus regani</i> (Ihering 1905)						11	5	5		21	3.0	33.3	2.3	77.77	8.8	MZUEL 09524	
<i>Hypostomus</i> cf. <i>strigaticeps</i> (Regan 1908)	267	77	93	210	119	36				802	6.0	66.7	89.1	5940.74	77.1	MZUEL 09366	
<i>Megalancistrus parananus</i> (Peters 1881)						5	5			10	2.0	22.2	1.1	24.69	5.0	MZUEL 09526	
<i>Pterygoplichthys ambrosetii</i> (Holmberg 1893) ♀						1	7			8	2.0	22.2	0.9	19.75	4.4	MZUEL 09555	

continue...

...continue

Táxon	Sampled Sections									Total	FO	FO%	Mean	FO%*Mean	KI	Voucher			
	A	B	C	D	E	F	G	H	I										
<b>Aspredinidae</b>																			
<i>Bunocephalus larai</i> Ihering 1930								3	1	4	2.0	22.2	0.4	9.87	3.1	MZUEL 09521			
<b>Auchenipteridae</b>																			
<i>Ageneiosus militaris</i> Valenciennes 1835									1	1	1.0	11.1	0.1	1.23	1.1	MZUEL 09459			
<i>Glanidium cesarpintoi</i> Ihering 1928									1	1	1.0	11.1	0.1	1.23	1.1	MZUEL 09539			
<i>Tatia neivai</i> (Ihering 1930)							1	1	3	11	2	18	5.0	55.6	2.0	111.11	10.5 MZUEL 09533		
<b>Doradidae</b>																			
<i>Rhinodoras dorbignyi</i> (Kner 1855)										13	13	1.0	11.1	1.4	16.04	4.0 MZUEL 09550			
<b>Heptapteridae</b>																			
<i>Cetopsorhamdia iheringi</i> Schubart & Gomes 1959									3		3	1.0	11.1	0.3	3.70	1.9 MZUEL 09483			
<i>Imparfinis mirini</i> Haseman 1911							1	2	13	3	1	20	5.0	55.6	2.2	123.45	11.1 MZUEL 09553		
<i>Imparfinis schubarti</i> (Gomes 1956)♦																			
<i>Phenacorhamdia tenebrosa</i> (Schubart 1964)										2		2	1.0	11.1	0.2	2.46	1.6 MZUEL 09484		
<i>Pimelodella meeki</i> Eigenmann 1910							1	3	8			12	3.0	33.3	1.3	44.44	6.7 MZUEL 09532		
<i>Rhamdia quelen</i> (Quoy & Gaimard 1824)							36	5	4	3	4	2	14	68	7.0	77.8	7.6	587.65	24.2 MZUEL 09548
<b>Pimelodidae</b>																			
<i>Iheringichthys labrosus</i> (Lütken 1874)							4	16	37	24	16	44	141	6.0	66.7	15.7	1044.44	32.3 MZUEL 09551	
<i>Megalonema platanum</i> (Günther 1880)										5		5	1.0	11.1	0.6	6.17	2.5 MZUEL 09474		
<i>Pimelodus maculatus</i> Lacépède 1803							1		14	8	28	51	4.0	44.4	5.7	251.85	15.9 MZUEL 09445		
<i>Pimelodus microstoma</i> Steindachner 1877							57	6	9	103	39	7	221	6.0	66.7	24.6	1637.03	40.5 MZUEL 09360	
<i>Pimelodus paranaensis</i> Britski & Langeani 1988										8	2	10	2.0	22.2	1.1	24.69	5.0 MZUEL 09438		
<i>Pinirampus pirinampu</i> (Spix & Agassiz 1829)										4		4	1.0	11.1	0.4	4.93	2.2 MZUEL 09475		
<i>Pseudoplatystoma corruscans</i> (Spix & Agassiz 1829)										1		1	1.0	11.1	0.1	1.23	1.1 MZUEL 09461		
<i>Sorubim lima</i> (Bloch & Schneider 1801) ♦										3		3	1.0	11.1	0.3	3.70	1.9 MZUEL 09462		
<b>Pseudopimelodidae</b>																			
<i>Rhyacoglanis paranensis</i> Shibatta & Vari 2017									115			115	1.0	11.1	12.8	141.97	11.9 MZUEL 14120		
<b>SYNBRANCHIFORMES</b>																			
<b>Synbranchidae</b>																			
<i>Synbranchus marmoratus</i> Bloch 1795									1			1	1.0	11.1	0.1	1.23	1.1 MZUEL 09439		
<b>CICHLIFORMES</b>																			
<b>Cichlidae</b>																			
<i>Australoheros tavaresi</i> Ottoni 2012							1					1	1.0	11.1	0.1	1.23	1.1 MZUEL 09447		
<i>Crenicichla britskii</i> Kullander 1982									4	9		13	2.0	22.2	1.4	32.09	5.7 MZUEL 09453		
<i>Crenicichla jaguarensis</i> Haseman 1911									7	8	22	37	3.0	33.3	4.1	137.03	11.7 MZUEL 09261		
<i>Crenicichla jupiaensis</i> Britski & Luengo 1968										1		1	1.0	11.1	0.1	1.23	1.1 MZUEL 09468		
<i>Geophagus iporangensis</i> (Haseman 1911)							11	182	238	183	3	10	627	6.0	66.7	69.7	4644.44	68.2 MZUEL 09353	
<i>Oreochromis niloticus</i> (Linnaeus 1758) ♦							2	1			3	7	1	14	5.0	55.6	1.6	86.41	9.3 MZUEL 09482

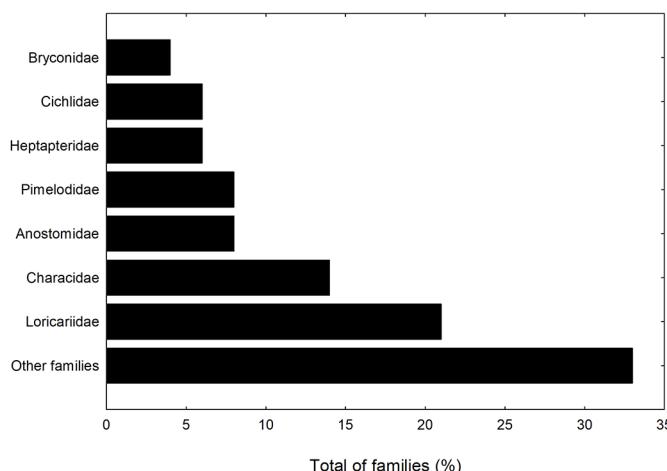
continue...

...continue

Táxon	Sampled Sections									Total	FO	FO%	Mean	FO%*Mean	KI	Voucher	
	A	B	C	D	E	F	G	H	I								
<b>CYPRINIDONTIFORMES</b>																	
<b>Poeciliidae</b>																	
<i>Phalloceros harpagos</i> Lucinda 2008	278		1043		27	3				1351	4.0	44.4	150.1	6671.60	81.7	MZUEL 09429	
<i>Poecilia reticulata</i> Peters 1859♦♣					5	5	15	31	38	38	57	60	64				
Number of species					0.49	0.42	0.54	0.12	0.1	0.07	0.06	0.08	0.05				
Dominance					0.49	0.52	0.17	0.39	0.39	0.49	0.43	0.34	0.46				
Evenness																	

## Results

A total of 11.934 specimens were collected, comprising seven orders, 27 families, and 100 species (Table 2; Appendix 1 - 5). Among these, the most representative order was Siluriformes (44.5%), followed by Characiformes (38.6%), Gymnotiformes (6.9%), and Cichliformes (5.9%). Cyprinodontiformes, Cypriniformes, and Synbranchiformes were represented by only one specimen each. Regarding the families, Loricariidae had 20 species (21%) and Characidae 16 species (14%). Together, these families showed the highest species richness (representing 35% of total richness), followed by Pimelodidae and Anostomidae, both composed of eight species each (8%), Heptapteridae and Cichlidae, both with six species each (6%), and Bryconidae with four species (4%; Figure 3).



**Figure 3.** Percentage composition of representative families of the ichthyofauna from the Laranjinha River, Upper Parana River basin, Paraná State, Brazil.

Among the sampling sites, A and B showed the lowest number of species (five species each), while sampling site I had a higher number of species (64 species). There is a progressive tendency of species richness from the source to the mouth ( $r^2 = 0.964$ ). The dominance (D) is higher in the upper Laranjinha River (sites A, B, and C showing D = 0.49, 0.42, and 0.54, respectively), and this dominance drops expressively on the sites D (0.12) until I (0.05).

The highest absolute abundance was observed in *Phalloceros harpagos* Lucinda 2008 (11.3%), *Hypostomus ancistroides* (Ihering 1911) (9.8%), *Hypostomus cf. paulinus* (Ihering 1905) (8.4%),

*Psalidodon aff. paranae* (6.9%), *Hypostomus cf. strigatus* (Regan 1908) (6.7%), and *Astyanax lacustris* (Lütken 1875) (5.3%; Table 2). Concerning the frequency of occurrence, the most frequent species of Laranjinha River were *Psalidodon aff. paranae* (FO = 88.8%), *Hoplias gr. malabaricus*, *Astyanax lacustris*, *Psalidodon bockmanni* (Vari & Castro 2007), *Rhamdia quelen* (Quoy & Gaimard 1824), *Hypostomus ancistroides*, *Hypostomus cf. paulinus*, and *Hypostomus topavae* (Godoy 1969) (FO = 77.7%). The classification of species from Kendigh abundance index highlights the following ranking: *Hypostomus ancistroides*, *Hypostomus cf. paulinus*, *Psalidodon aff. paranae*, *Phalloceros harpagos* Lucinda 2008, *Hypostomus cf. strigatus*, and *Astyanax lacustris*. Among these species, *Psalidodon aff. paranae* and *Phalloceros harpagos* were frequent and dominant in the headwater region (sites A, B, and C), while the others were frequent downstream those sites.

From all recorded species, 9.8% were non-native: *Aphyocharax cf. dentatus* Eigenmann & Kennedy 1903, *Apterodonotus cf. caudimaculosus* Santana 2003, *Cyprinus carpio* Linnaeus 1758, *Hyphessobrycon eques* (Steindachner 1882), *Loricariichthys platymetopon* Isbrücker & Nijssen 1979, *Metynnis lippincottianus* (Cope 1870), *Oreochromis niloticus* (Linnaeus 1758), *Poecilia reticulata* Peters 1859, *Pterygoplichthys amboinensis* (Holmberg 1893), *Sorubim lima* (Bloch & Schneider 1801), and *Triportheus nematurus* (Kner, 1858). Of these species, two were exotic: *Cyprinus carpio*, which is a carp from Asia, and *Oreochromis niloticus*, known as Nile-tilapia, from Africa.

Ten long-distance migratory species were recorded in the Laranjinha River basin: *Brycon orbygnianus* (Valenciennes 1850), *Megaleporinus obtusidens* (Valenciennes 1837), *Piaractus mesopotamicus* (Holmberg 1887), *Pinirampus pirinampu* (Spix & Agassiz 1829), *Prochilodus lineatus* (Valenciennes 1837), *Pseudoplatystoma corruscans* (Spix & Agassiz 1829), *Rhinelepis aspera* Spix & Agassiz 1829, *Salminus brasiliensis* (Cuvier 1816), and *Salminus hilarii* Valenciennes 1850. Additionally, *Leporinus friderici* (Bloch 1794), *Pimelodus maculatus* Lacepede 1803, *Rhamdia quelen*, and *Schizodon nasutus* Kner 1858, which are considered short-migratory species, were also recorded herein (Agostinho et al. 2007, Oliveira et al. 2015).

Among the collected species, *Apterodonotus acidops* Triques 2011, *Brycon orbygnianus*, *Brycon nattereri* Günther 1864, *Crenicichla jupiaensis* Britski & Luengo 1968, and *Rhinelepis aspera* Spix & Agassiz 1829 are classified as endangered in the most recent IUCN Red List. It is worth mentioning that two species are possibly new to science, *Piabarchus aff. stramineus* (sensu Frota et al., 2016), and *Eigenmannia* sp.

## Discussion

The results presented in this study showed a high diversity of fishes (100 species) among the fish collected in the Laranjinha River. Such results follow the pattern found in the Neotropical regions, showing the dominance of Siluriformes and Characiformes (Agostinho et al. 1997, Lowe-McConnell 1999, Jarduli et al. 2020). Furthermore, among Paranapanema River tributaries, the number of species is lower if compared to the Tibagi River basin, where 158 species were registered (Lobón-Cerviá & Bennemann 2000, Bennemann et al. 2006, 2011, Shibatta & Cheida 2003, Hoffmann et al. 2005, Oliveira & Bennemann et al. 2005, Jerep et al. 2006, Sant'Anna et al. 2006, Shibatta et al. 2002, 2006a, 2006b, 2007, 2008, Galves et al. 2007, Vieira & Shibatta 2007, Orsi 2010, Raio & Bennemann 2010, Garcia et al. 2014, 2015, Silva et al. 2015, Frantine-Silva et al. 2015, Almeida et al. 2018, Jerep & Shibatta 2017, Claro-Garcia et al. 2018), and to the Cinzas River, with 114 species (Hoffmann et al. 2005, Vianna & Nogueira, 2008, Orsi 2010, Bennemann et al. 2011, Costa et al. 2013, Frantine-Silva et al. 2015, Almeida et al. 2018).

It is noteworthy that the most frequent species of Laranjinha River was *Psalidodon* aff. *paranae*, captured in eight sites. The higher abundance were in the uppermost site A, and along the downstream, the number of specimens decreased until site H, which corroborate the hypothesis that this species has a preference regarding the headwater region (Britski 1972), but it is not restricted to that. *Psalidodon* aff. *paranae* is an insectivorous species, feeding mainly of allochthonous resources from the riparian forest. However, it can gather enough food resources from environments with different levels of degradation (Ferreira et al. 2012).

In the upper region of Laranjinha River, *Phalloceros harpagos*, a native species, was the most abundant one, being dominant in the site C. The value of dominance index ( $D=0.54$ ) and evenness ( $E=0.17$ ) of that site, reflected the disturbance in the observed environment. Biological features of *P. harpagos* like constant reproduction (Wolff et al. 2007), adaptability, tolerance to heat, variations in salinity (Nascimento & Gurgel 2000), and high trophic plasticity (Casatti et al. 2009, Rocha et al. 2009) may be related to the success of the species at that location. This species has a wide geographical distribution (Thomaz et al. 2019), which may also be related to its ecological plasticity.

*Hypostomus ancistroides*, *H. cf. paulinus*, and *H. cf. strigaticeps* were abundant and frequent from C to I sites. A factor that possibly has favored these rheophilic species is the presence of running water in the Laranjinha River (Cecilio et al. 1997; Garcia et al. 2020). Several species of *Hypostomus* were collected in running waters showing a substrate with pebbles and rocks (Garavello & Garavello 2004; Perez-Junior & Garavello 2007), but in Laranjinha River these species also occur in some places with a sand bottom. *Hypostomus ancistroides*, the most abundant of these species, had higher abundance in the site C, which decreased until site I, and such data demonstrates that this species is the least rheophilic one among congeners.

In general, the non-native fish species recorded in this study were introduced from other drainages in Brazil. All these species are associated with human activities like aquarium trade (possibly *Aphyocharax* cf. *dentatus*, *Apteronotus albifrons*, *Hyphessobrycon eques* and *Poecilia reticulata*), fish ladders (possibly *Metynnis lippincottianus*, *Pterygoplichthys ambrosetti* and *Triportheus nematurus*), fish farming (certainly *Cyprinus carpio*, *Oreochromis niloticus* and *Sorubim lima*),

and control of insect larvae (possibly *Poecilia reticulata*). It is worth mentioning that among these non-native species, two of them are considered exotic (*Cyprinus carpio* and *Oreochromis niloticus*). These records are alarming, as *C. carpio* is known to promote bioturbation by continually revolving the sediment (Ritvo et al. 2004). However, only two individuals were sampled, being exclusive on site G, an area surrounded by pastures and agriculture. On the other hand, *O. niloticus* was more abundant and widely distributed, being sampled in five sites, which corresponds to an area surrounded by pasture and agriculture, and specifically in site B with the discharge of effluents. In addition, this species changes the environment due to the excess of nitrogen excreta, thus favoring the proliferation of algae, which decreases light and dissolved oxygen (Britton et al. 2007, Vicente & Fonseca-Alves 2013). Besides that, *Poecilia reticulata* was registered by Costa et al. (2013) in the study of the Penacho stream, a small tributary on the right bank of the Laranjinha River that flows into the Cinzas River. The high abundance of *P. reticulata* indicates an instability of the environment, including factors such as the lack of food resources for other species and the low level of dissolved oxygen (De Souza & Tozzo 2013).

The occurrence of 13 medium- to large-size migratory fishes highlights the importance of Laranjinha River for the maintenance of the diversity and viability of the ichthyofauna of the middle Paranapanema River. Other studies also highlight the Cinzas River basin and the Laranjinha River as migratory routes to the fish fauna from the Capivara dam (Dias et al. 2004, Lopes et al. 2007, Vianna & Nogueira 2008, Orsi 2010). As the Laranjinha River has only a little dam, including a fish pass system built in it, it extends a long stretch without dams, enabling migratory fish species to use its free-flowing stretches to complete their reproductive cycles. Long-distance migratory species with high commercial value have been using tributaries of rivers intensely affected by dams as migratory routes (Agostinho et al. 2008). The pressure caused by power plant dam constructions is intense in freshwater systems (Agostinho et al. 2005), because the transformation of lotic areas into lentic ones interrupts the displacement of migratory fish (Agostinho et al. 2008, Pelicice et al. 2018). The impact is not restricted to migratory species, and it also impairs the local fauna that depends on the tributaries for the viability of spawning and survival of early life stages (Oliveira et al. 2015). Studying the dynamics of eggs and larvae in the Cinzas River, Vianna & Nogueira (2008), found that the fishes of the middle Paranapanema River use this tributary to spawn. In general, biological communities are under heavy pressure due to environmental instability caused by habitat fragmentation and loss of natural environments, which affect species abundance and richness (Pusey & Arthington 2003, Di Giulio et al. 2009, Shandas & Alberti, 2009).

Furthermore, the conservation of the Laranjinha River is essential to preserve the species already categorized as threatened by extinction in the IUCN's Red List (*Apteronotus acidops*, *Brycon orbygnianus* and *Crenicichla jupiaensis* as Endangered; *Brycon nattereri* as Vulnerable; and *Rhinelepis aspera* as Near Threatened) (ICMBio 2018). The fact that some of these fish appear on a list of endangered species is concerning and it reinforces that the focused area needs appropriate conservation strategies (Simic et al. 2007). Studies on *B. nattereri* reinforce the importance of Laranjinha River conservation, since this migratory and threatened species has been able to maintain a satisfactory population genetic diversity due to the quality of that environment (Galindo et al. 2019).

Finally, the Laranjinha River is a heritage of fish diversity due to the presence of large numbers of species (among migratory, threatened, and new species to science). Thus, it deserves special attention regarding its preservation.

## Supplementary Material

The following online material is available for this article:

[Appendix 1](#)

[Appendix 2](#)

[Appendix 3](#)

[Appendix 4](#)

[Appendix 5](#)

## Acknowledgments

Our thanks to the “ Universidade Estadual do Norte do Paraná - UENP/PROPG/EDITORA UENP” for the partial support provided

## Author Contributions

Bruno Ambrozio Galindo: Substantial contribution in the concept and design of the study; Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Renata Rúbia Ota: Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Thiago Deruza Garcia: Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Raul Henrique Cardoso do Nascimento: Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation.

Willian Massaharu Ohara: Contribution to data collection; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Augusto Seawright Zanatta: Contribution to data collection; Contribution to manuscript preparation.

Dhiego Gomes Ferreira: Substantial contribution in the concept and design of the study; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Caroline Apolinário-Silva: Substantial contribution in the concept and design of the study; Contribution to data collection; Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation.

Wilson Frantine-Silva: Contribution to data collection; Contribution to manuscript preparation.

Sandremir de Carvalho: Substantial contribution in the concept and design of the study; Contribution to data analysis and interpretation; Contribution to manuscript preparation.

Alexandro Derly Augusto Costa: Contribution to data collection; Contribution to manuscript preparation.

Silvia Helena Sofia: Substantial contribution in the concept and design of the study; Contribution to data analysis and interpretation; Contribution to manuscript preparation.

Oscar Akio Shibatta: Substantial contribution in the concept and design of the study; Contribution to data analysis and interpretation; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

## Conflicts of Interest

The authors declare that they have no conflict of interest related to the publication of this manuscript.

## References

- AGOSTINHO, A.A., GOMES, L.C. & PELICICE, F.M. 2007. Ecology and management of fish resources in reservoirs in Brazil. Eduem.
- AGOSTINHO, A.A., JÚLIO-JR, H.F., GOMES, L.C., BINI, L.M. & AGOSTINHO, C.S. 1997. Composição, abundância e distribuição espaço-temporal da ictiofauna, p. 179-208. In: VAZZOLER, A.E.A.M., AGOSTINHO, A.A. & HAHN, N.S. (Eds.). A planície de inundação do alto rio Paraná: aspectos físicos, biológicos e socioeconômicos. EDUEM. p. 460.
- AGOSTINHO, A.A., GOMES, L.C., FERNANDES, D.R. & SUZUKI, H.I. 2002. Efficiency of fish ladders for neotropical ichthyofauna. River Research and Applications. 18(3):299–306.
- AGOSTINHO, A.A., PELICICE, F.M. & GOMES, L.C. 2008. Dams and the fish fauna of the Neotropical region: impacts and management related to diversity and fisheries. Brazilian Journal of Biology. 68(4):1119-1132.
- AGOSTINHO, A.A., THOMAZ, S.M. & GOMES, L.C. 2005. Conservation of the biodiversity of Brazil's inland waters. Conservation Biology. 19(3):646-652.
- ALMEIDA, F.S., FRANTINE-SILVA, W., LIMA, S.C., GARCIA, D.A.Z. & ORSI, M.L. 2018. DNA barcoding as a useful tool for identifying non-native species of freshwater ichthyoplankton in the neotropics. Hydrobiologia. 817(1):111-119.
- AZEVEDO-SANTOS, V.M., VITULE, J.R.S., GARCÍA-BERTHOU, E., PELICICE, F.M. & SIMBERLOFF, D. 2016. Misguided strategy for mosquito control. Science. 351:675.
- BENNEMANN, S.T., Capra, L.G., Galves, W. & Shibatta, O.A. 2006. Dinâmica trófica de *Plagioscion squamosissimus* (Perciformes, Sciaenidae) em trechos de influência da represa Capivara (rios Paranapanema e Tibagi). Iheringia. Série Zoologia. 96(1):115-119.
- BENNEMANN, S.T., GALVES, W. & CAPRA, L.G. 2011. Recursos alimentares utilizados pelos peixes e estrutura trófica de quatro trechos no reservatório Capivara (Rio Paranapanema). Biota Neotropica. 11(1):63-71.
- BRITSKI, H.A. 1972. Peixes de água doce do Estado de São Paulo, p. 79-108. In: Comissão Interestadual da Bacia Paraná-Uruguai, Poluição e piscicultura. Sistemática. Faculdade de Saúde Pública USP e Instituto de Pesca.
- BRITTO, S.G.C., SIROL, R.N., VIANNA, N.C., JARDIM, S.M., SANTOS, J.C. & PELISARI, E. 2003. Peixes do rio Paranapanema. São Paulo: Duke Energy Internacional Geração Paranapanema. p.112.
- BRITTON, J.R., BOAR, R.R., GREY, J., FOSTER, J., LUGONZO, J. & HARPER, D.M. 2007. From introduction to fishery dominance: the initial impacts of the invasive carp *Cyprinus carpio* in Lake Naivasha, Kenya, 1999 to 2006. Journal of Fish Biology. 71:239-257.
- BRITTON, J.R. & ORSI, M.L. 2012. Non-native fish in aquaculture and sport fishing in Brazil: economic benefits versus risks to fish diversity in the upper River Paraná Basin. Reviews in Fish Biology and Fisheries. 22(3):555–565.
- BRITTO, S.G.C. & CARVALHO, E.D. 2006. Ecological attributes of fish fauna in the Taquarucu Reservoir, Paranapanema River (Upper Paraná, Brazil): composition and spatial distribution. Acta Limnologica Brasiliensis. 18(4):377–388.

- CASATTI, L., FERREIRA, C.P. & LANGEANI, F. 2009. A fish-based biotic integrity index for assessment of lowland streams in southeastern Brazil. *Hydrobiologia*. 623:173-189.
- CASTRO, R.M.C., CASATTI, L., SANTOS, H.F., FERREIRA, K.M., RIBEIRO, A.C., BENINE, R.C., DARDIS, G.Z.P., MELO, A.L.A., STOPIGLIA, R., ABREU, T.X., BOCKMANN, F.A., CARVALHO, M., GIBRAN, F.Z. & LIMA, F.C.T. 2003. Estrutura e composição da ictiofauna de riachos do rio Paranapanema, sudeste e sul do Brasil. *Biota Neotropica*. 3(1):1-31.
- CAVALLI, D., FROTA, A., LIRA, A.D., GUBIANI, É.A., MARGARIDO, V.P. & GRAÇA, W.J.D. 2018. Update on the ichthyofauna of the Piquiri River basin, Paraná, Brazil: a conservation priority area. *Biota Neotropica*. 18(2):e20170350.
- CECILIO, E.B., AGOSTINHO, A.A., JÚLIO-JÚNIOR, H.F. & PAVANELLI, C.S. 1997. Colonização ictiofaunística do reservatório de Itaipu e áreas adjacentes. *Revista Brasileira de Zoologia*. 14:1-14.
- CETRA, M., BARRELLA, W., NETO, F.L., MARTINS, A.G., MELLO, B.J. & ALMEIDA, R.S. 2012. Fish fauna of headwater streams that cross the Atlantic Forest of south São Paulo state. *Check List*. 8(3):421-425.
- CETRA, M., MATTOX, G.M.T., FERREIRA, F.C., GUINATO, R.B., SILVA, F.V. & PEDROSA, M. 2016. Headwater stream fish fauna from the Upper Paranapanema River basin. *Biota Neotropica*. 16(3): e20150145.
- CIONEK, V.M., SACRAMENTO, P.A., ZANATTA, N., OTA, R.P., CORBETTA, D.F. & BENEDITO, E. 2012. Fishes from first order streams of lower Paranapanema and Ivaí rivers, upper Paraná River basin, Paraná, Brazil. *Check List*. 8(6):1158-1162.
- CLARO-GARCÍA, A., ASSEGÁ, F.M. & SHIBATTA, O.A. 2018. Diversity and distribution of ichthyofauna in streams of the middle and lower Tibagi river basin, Paraná, Brazil. *Check List*. 14:43.
- COSTA, A.D.A., FERREIRA, D.G., SILVA, W.F., ZANATTA, A.S., SHIBATTA, O.A., & GALINDO, B.A. 2013. Fishes (Osteichthyes: Actinopterygii) from the Penacho stream, upper Paraná River basin, Paraná State, Brazil. *Check List*. 9(3):519-523.
- COSTELLO, D.M., TIEGS, S.D., & LAMBERTI, G.A. 2011. Do non-native earthworms in Southeast Alaska use streams as invasional corridors in watersheds harvested for timber? *Biological Invasions*. 13(1):177-187.
- DE SOUZA, F. & TOZZO, R. A. 2013. *Poecilia reticulata* Peters 1859 (Cyprinodontiformes, Poeciliidae) como possível bioindicador de ambientes degradados. *Revista Meio Ambiente e Sustentabilidade*. 3(2):162-175.
- DI GIULIO, M., HOLDERECKER, R. & TOBIAS, S. 2009. Effects of habitat and landscape fragmentation on humans and biodiversity in densely populated landscapes. *Journal of environmental management*. 90(10):2959-2968.
- DIAS, J.H., BRITTO, S.G.C., VIANNA, N.C. & GARAVELLO, J.C. 2004. Biological and ecological aspects of *Pinirampus pirinampu* (Spix, 1829) Siluriformes, Pimelodidae. Capivara reservoir, Paranapanema River Southern Brazil. *Acta Limnologica Brasileira* 16:293-304.
- FERREIRA, A., GERHARD, P. & CYRINO, J.E.P. 2012. Diet of *Astyanax paranae* (Characidae) in streams with different riparian land covers in the Passa Cinco River basin, southern Brazil. *Iheringia, Série Zoologia* 10:80-87.
- FRANTINE-SILVA, W., FERREIRA, D.G., NASCIMENTO, R.H.C., FRACASSO, J.F., CONTE, J.E., RAMOS, F.P., CARVALHO, S. & GALINDO, B.A. 2015. Genetic analysis of five sedentary fish species in middle Laranjinha River (upper Paraná River basin): A case study. *Genetics and Molecular Research*. 14(4):18637-18649.
- FRICKE, R., ESCHMEYER, W.N. & VAN DER LAAN, R. 2020. Catalog of fishes: genera, species, references. Disponível em: (<http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>). Access: March 23<sup>th</sup>, 2020.
- FROEHLICH, O., CAVALLARO, M., SABINO, J., SÚAREZ, Y.R., & VILELA, M.J.A. 2017. Checklist da ictiofauna do estado de Mato Grosso do Sul, Brasil. *Iheringia, Série Zoologia*. 107: e2017151.
- FROTA, A., DEPRÁ, G.D.C., PETENUCCI, L.M. & GRAÇA, W.J. 2016. Inventory of the fish fauna from Ivaí River basin, Paraná State, Brazil. *Biota Neotropica*. 16(3): e20150151.
- FROTA, A., OLIVEIRA, R.C.D., BENEDITO, E. & GRAÇA, W.J. 2019. Ichthyofauna of headwater streams from the rio Ribeira de Iguape basin, at the boundaries of the Ponta Grossa Arch, Paraná, Brazil. *Biota Neotropica*. 19(1):e20180666.
- FROTA, A., OTA, R.R., DEPRÁ, G.C., GANASSIN, M.J.M & GRAÇA, W.J. 2020. A new inventory for fishes of headwater streams from the rio das Cinzas and rio Itararé basins, rio Paranapanema system, Paraná, Brazil. *Biota Neotropica*. 20(1): e20190833.
- GALINDO, B.A., FERREIRA, D.G., APOLINÁRIO-SILVA, C., TERRA, M.C., APRÍGIO, N.G., OTA, R.R., OHARA, W.M., ALMEIDA, F.S. & SOFIA, S.H. 2019. Genetic diversity and population structure of *Brycon nattereri* (Characiformes: Bryconidae): a Neotropical fish under threat of extinction. *Neotropical Ichthyology*. 17(1):e180071.
- GALVES, W., SHIBATTA, O.A. & JEREPE, F.C. 2007. Fish, Taquara river basin, northern of the state of Paraná, Brazil. *Check List*. 3(3):253-259.
- GARAVELLO, J.C. & GARAVELLO, J.P. 2004. Spatial distribution and interaction of four species of the catfish genus *Hypostomus* Lacépède with bottom of Rio São Francisco, Canindé do São Francisco, Sergipe, Brazil (Pisces, Loricariidae, Hypostominae). *Brazilian Journal of Biology*. 64(3B):103-141.
- GARCIA, D.A.Z., COSTA, A.D.A., LEME, G.L.A. & ORSI, M.L. 2014. Biology of black bass *Micropterus salmoides* (Lacépède, 1802) fifty years after the introduction in a small drainage of the Upper Paraná River basin, Brazil. *Biodiversitas*. 15(2):180-185.
- GARCIA, D.A.Z., HERNANDES, M.C., SILVA-SOUZA, Â.T. & ORSI, M.L. 2015. Establishment of non-native predator (Pisces, Erythrinidae) in a tributary of the Upper Paraná River basin, south Brazil. *Neotrop. Biol. Conserv.* 10(3):177-181.
- GARCIA, T.D., OTA, R.R., FERREIRA, D.G., NASCIMENTO, R.H., GALINDO, B.A., PEREIRA, L.S., & ZANATTA, A.S. 2020. Distribution of Siluriformes in a river under the influence of a small hydroelectric power plant of the Paraná River Basin, Brazil. *Iheringia, Série Zoologia*. 110: e2020005.
- HAMMER, M., YAROSLAVSKY, A.N. & SCHWEITZER, D. 2001. A scattering phase function for blood with physiological haematocrit. *Physics in Medicine & Biology*. 46(3):65.
- HOFFMANN, A.C., ORSI, M.L. & SHIBATTA, O.A. 2005. Diversidade de peixes do reservatório da UHE Escola Engenharia Mackenzie (Capivara), rio Paranapanema, bacia do alto rio Paraná, Brasil, e a importância dos grandes tributários na sua manutenção. *Iheringia, Série Zoológica*. 95(3):319-325.
- ICMBio 2018. Red Book of Threatened Brazilian Fauna. Disponível em: <<https://www.nationalredlist.org/livro-vermelho-da-fauna-brasileira-ameaçada-de-extinção-2018-red-book-of-threatened-brazilian-fauna-portuguese>>. December 18<sup>th</sup>, 2019.
- JARDULI, L. R., GARCIA, D. A. Z., VIDOTTO-MAGNONI, A. P., CASIMIRO, A. C. R., VIANNA, N. C., ALMEIDA, F. S. D., JEREPE, F. C. & ORSI, M. L. 2020. Fish fauna from the Paranapanema River basin, Brazil. *Biota Neotropica*. 20(1).
- JEREPE, F.C. & SHIBATTA, O.A. 2017. A new species of *Bryconamericus* (Characidae: Stevardiinae: Diapomini) from the upper rio Paraná basin, Brazil. *Neotropical Ichthyology*. 15(3):e170028.
- JEREPE, F.C., SHIBATTA, O.A., PEREIRA, E.H. & OYAKAWA, O.T. 2006. Two new species of *Ishbrueckerichthys* Derijst, 1996 (Siluriformes: Loricariidae) from the rio Paranapanema basin, Brazil. *Zootaxa*. 1372(1):53-68.
- JÚLIO-JÚNIOR, H.F., TÓS, C.D., AGOSTINHO, A.A. & PAVANELLI, C.S. 2009. A massive invasion of fish species after eliminating a natural barrier in the upper rio Paraná basin. *Neotropical Ichthyology*. 7(4):709-718.
- LANGEANI, F., CASTRO, R.M.C., OYAKAWA, O.T., SHIBATTA, O.A., PAVANELLI, C.S. & CASATTI, L. 2007. Diversidade da ictiofauna do Alto Rio Paraná: composição atual e perspectivas futuras. *Biota Neotropica*. 7(3):1-17.
- LANSAC-TÔHA, F.M., HEINO, J., QUIRINO, B.A., MORESCO, G.A., PELÁEZ, O., MEIRA, B.R., RODRIGUES, L.C., JATI, S., LANSAC-TÔHA, F.A. & VELHO, L. F. M. 2019. Differently dispersing organism groups show contrasting beta diversity patterns in a dammed subtropical river basin. *Science of The Total Environment*, 691, 1271-1281.
- LOBÓN-CERVÍA, J. & BENNEMANN, S. 2000. Temporal trophic shifts and feeding diversity in two sympatric, neotropical, omnivorous fishes: *Astyanax bimaculatus* and *Pimelodus maculatus* in Rio Tibagi (Paraná, Southern Brazil). *Archiv für Hydrobiologie*. 285-306.

## Ichthyofauna of the Laranjinha River basin

- LOPES, C.M., ALMEIDA, F.S., ORSI, M.L., BRITTO, S.G.D.C., SIROL, R.N. & SODRÉ, L.M.K. 2007. Fish passage ladders from Canoas Complex-Paranapanema River: evaluation of genetic structure maintenance of *Salminus brasiliensis* (Teleostei: Characiformes). *Neotropical Ichthyology*. 5(2):131-138.
- MATTSON, N.S. & RIPPLE, T.H. 1989. Metomidate, a better anesthetic for cod (*Gadus morhua*) in comparison with benzocaine, MS-222, chlorobutanol, and phenoxyethanol. *Aquaculture*. 83(1-2):89-94.
- NASCIMENTO, R.S.S. & GURGEL, H.D.C.B. 2000. Estrutura populacional de *Poecilia vivipara* Bloch & Schneider, 1801 (Atheriniformes, Poeciliidae) do rio Ceará-Mirim-Rio Grande do Norte. *Acta Scientiarum. Biological Sciences*. 22:415-422.
- OLIVEIRA, D.C.D., & BENNEMANN, S.T. 2005. Ictiofauna, recursos alimentares e relações com as interferências antrópicas em um riacho urbano no sul do Brasil. *Biota Neotropica*. 5(1):95-107.
- OLIVEIRA, A.G., SUZUKI, H.I., GOMES, L.C. & AGOSTINHO, A.A. 2015. Interspecific variation in migratory fish recruitment in the Upper Paraná River: effects of the duration and timing of floods. *Environmental Biology of Fishes*. 98(5):1327-1337.
- ORSI, M.L. 2010. Estratégias reprodutivas de peixes da Região média-baixa do Rio Paranapanema, Reservatório de Capivara. São Paulo. Blucher Acadêmico. p.115.
- ORSI, M.L. & AGOSTINHO, A.A. 1999. Introdução de espécies de peixes por escapes acidentais de tanques de cultivo em rios da Bacia do Rio Paraná, Brasil. *Revista brasileira de Zoologia*. 16(2):557-560.
- ORTEGA, J.C.G., JÚLIO JR, H.F., GOMES, L.C. & AGOSTINHO, A.A. 2015. Fish farming as the main driver of fish introductions in Neotropical reservoirs. *Hydrobiologia*. 746:147-158.
- OTA, R.R., DA GRAÇA, W.J., & PAVANELLI, C.S. 2015. Neotropical Siluriformes as a model for insights on determining biodiversity of animal groups. *PloS one*. 10(7):e0132913.
- OTA, R.R., DEPRÁ, G.D.C., GRAÇA, W.J.D. & PAVANELLI, C.S. 2018. Peixes da planície de inundação do alto rio Paraná e áreas adjacentes: revised, annotated and updated. *Neotropical Ichthyology*. 16(2):e170094[1]-e170094[11].
- PELICICE, F. M., AZEVEDO-SANTOS, V.M., ESGUÍCERO, A.L.H., AGOSTINHO, A.A. & ARCIFA, M.S. 2018. Fish diversity in the cascade of reservoirs along the Paranapanema River, southeast Brazil. *Neotrop. Ichthyol.* 16(2):e170150.
- PEREZ-JUNIOR, O.R. & GARAVELLO, J.C. 2007. Ictiofauna do ribeirão do Pântano, afluente do rio Mogi-Guaçu, bacia do alto rio Paraná, São Paulo, Brasil. *Iheringia. Série Zoologia*. 97(3):328-335.
- PUSEY, B.J. & ARTHINGTON, A.H. 2003. Importance of the riparian zone to the conservation and management of freshwater fish: a review. *Marine and freshwater Research*. 54(1):1-16.
- RAIO, C.B. & BENNEMANN, S.T. 2010. A ictiofauna da bacia do rio Tibagi e o projeto de construção da UHE Mauá, Paraná, Brasil. *Semina: Ciências Biológicas e da Saúde*. 31(1):15-20.
- RITVO, G., KOCHBA, M. & AVNIMELECH, Y. 2004. The effects of common carp bioturbation on fishpond bottom soil. *Aquaculture*. 242(4):345-356.
- ROCHA, F.C., CASATTI, L. & PEREIRA, D.C. 2009. Structure and feeding of a stream fish assemblage in Southeastern Brazil: evidence of low seasonal influences. *Acta Limnologica Brasiliensia*. 21(1):123-134.
- SANT'ANNA, J.F.M., ALMEIDA, M.C., VICARI, M.R., SHIBATTA, O.A. & ARTONI, R.F. 2006. Levantamento rápido de peixes em uma lagoa marginal do rio Imbituba na bacia do Alto rio Tibagi, Paraná, Brasil. *Publicatio. UEPG Ciências Biológicas e da Saúde*. 12(1):39-46.
- SANTOS, L.J.C., OKA-FIORI, C., CANALI, N.E., FIORI, A.P., SILVEIRA, C.T.D., SILVA, J.M.F.D. & ROSS, J.L.S. 2006. Mapeamento geomorfológico do Estado do Paraná. *Revista Brasileira de Geomorfologia*. 7:03-12.
- SCHWARTZ, W. 2006. Meio Ambiente Finalmente, a escada de peixes no Laranjinha Demorou sete anos para ficar pronta e será "inaugurada" na piracema que se aproxima. *Folha de Londrina*. Outubro, 27 de 2006.
- SHANDAS, V. & ALBERTI, M. 2009. Exploring the role of vegetation fragmentation on aquatic conditions: Linking upland with riparian areas in Puget Sound lowland streams. *Landscape and Urban Planning*. 90(1-2):66-75.
- SHIBATTA, O.A., BENNEMANN, S.T., MORI, H. & SILVA, D.F. 2008. Riqueza biológica e ecológica dos peixes do Ribeirão Varanal. In: *A flora e a fauna do Ribeirão Varanal: um estudo da biodiversidade do Paraná* (S.T. Bennemann, O.A. Shibatta, A.O.D. Vieira, eds) Londrina, EDUEL, p.77-97.
- SHIBATTA, O.A. & CHEIDA, C.C. 2003. Composição em tamanho dos peixes (Actinopterygii, Teleostei) de ribeirões da bacia do rio Tibagi, Paraná, Brasil. *Revista Brasileira de Zoologia*. 20(3):469-473.
- SHIBATTA, O.A., GEALH, A.M. & BENNEMANN, S.T. 2007. Ictiofauna dos trechos alto e médio da bacia do rio Tibagi, Paraná, Brasil. *Biota Neotropica*. 7:125-134.
- SHIBATTA, O.A., ORSI, M.L., BENNEMANN, S.T. & SILVA-SOUZA, A.T. 2002. Diversidade e distribuição de peixes na bacia do rio Tibagi; p.403-423, in: MEDRI, M.E., BIANCHINI, E., SHIBATTA, O.A. & PIMENTA, J.A. *A bacia do rio Tibagi*. Londrina.
- SHIBATTA, A.O., ORSI, M.L. & ARTONI, R.F. 2006a. Estratégia reprodutiva dos peixes do Parque Estadual de Vila Velha. In: *Peixes do Parque Estadual de Vila Velha: aspectos da história natural, da biologia evolutiva e da conservação* (R.F. Artoni & O.A. Shibatta, eds). Editora UEPG, Ponta Grossa, p.67-77.
- SHIBATTA, O.A., ORSI, M.L. & BENNEMANN, S.T. 2006b. Os peixes do Parque Estadual Mata dos Godoy. In: *Ecologia do Parque Estadual Mata do Godoy* (J.M. Torezan, org). Ed. Itedes, Londrina. p.156-167.
- SILVA, J.C., ROSA, R.R., GALDIOLI, E.M., SOARES, C.M., DOMINGUES, W.M., VERÍSSIMO, S. & BIALETZKI, A. 2017. Importance of dam-free stretches for fish reproduction: the last remnant in the Upper Paraná River. *Acta Limnologica Brasiliensis*, 29:e16.
- SIMIC, V., SIMIC, S., PAUNOVIC, M. & CAKIC, P. 2007. Model of the assessment of the critical risk of extinction and the priorities of protection of endangered aquatic species at the national level. *Biodiversity Conservation*. 16:2471-2493.
- STATSOFT, Inc. 2011. STATISTICA (data analysis software system).
- TERÁN, G. E., BENITEZ, M. F. & MIRANDE, J. M. 2020. Opening the Trojan horse: phylogeny of *Astyanax*, two new genera and resurrection of *Psalidodon* (Teleostei: Characidae). *Zoological Journal of the Linnean Society*. p. 1-18.
- TYTLER, P. & HAWKINS, A.D. 1981. Vivisection, anaesthetics and minor surgery. In: Hawkins, A.D. (Editor). *Aquarium Systems*. Academic Press. p. 247-278.
- THOMAZ, A. T., CARVALHO, T. P., MALABARBA, L. R. & KNOWLES, L. L. 2019. Geographic distributions, phenotypes, and phylogenetic relationships of *Phalloceros* (Cyprinodontiformes: Poeciliidae): Insights about diversification among sympatric species pools. *Molecular phylogenetics and Evolution*. 132:265-274.
- VAN DER LAAN, R., ESCHMEYER, W. N. & FRICKE, R. 2020. Family-group names of recent fishes. *Zootaxa*. 3882(1)230. Disponível em: (<https://www.calacademy.org/scientists/catalog-of-fishes-classification/>). Access: March 23<sup>th</sup>, 2020.
- VIANNA, N.C. & NOGUEIRA, M.G. 2008. Ichthyoplankton and limnological factors in the Cinzas River – an alternative spawning site for fishes in the middle Paranapanema River basin, Brazil. *Acta Limnologica Brasiliensis*. 20:139-151.
- VIEIRA, D.B. & SHIBATTA, O.A. 2007. Peixes como indicadores da qualidade ambiental do ribeirão Esperança, município de Londrina, Paraná, Brasil. *Biota Neotropica*. 7(1):bn01407012007.
- VICENTE, I.S.T. & FONSECA-ALVES, C.E. 2013. Impact of introduced Nile tilapia (*Oreochromis niloticus*) on non-native aquatic ecosystems. *Pakistan Journal of Biological Sciences*. 16(3):121-126.
- WOLFF, L.L., ERICSSON, H.R., VIANA, D. & ZALESKI, D. 2007. Population structure of *Phalloceros caudimaculatus* (Hensel, 1868) (Cyprinodontiformes, Poeciliidae) collected in a brook in Guarapuava, PR. *Brazilian Archives of Biology and Technology*. 50(3):417-423.

*Received: 21/01/2020**Revised: 15/08/2020**Accepted: 19/08/2020**Published online: 25/08/2020*